

Agar

Cat. # P1110-500, P1110-2000, P1110-5000

Also Known as:	N/A
Form:	Powder
Quantity:	See tube label
Technical Specification	Gel strength (1.5%): $\geq 800\text{g/cm}^2$ Moisture: $<15\%$ Ash: $\leq 2\%$ Gelation point: $32-39\text{ }^\circ\text{C}$
Description:	As a solidifying agent, thickening agent, mainly as a solidifying agent for preparing microbiological culture media.
Storage:	Ship under ambient. Keep container tightly closed and store at a dry place under room temperature.
Usage	A general protocol for preparing microbiological culture plate using agar. Preparing LB/agar medium <ol style="list-style-type: none">1. Weigh out 25 g of granular LB (Miller's) (UBPBio P1310) or: 10 g peptone (UBPBio P1210) + 5 g yeast extract (UBPBio P1220) + 10 g NaCl2. Weigh out 7.5 g agar (UBPBio P1110)3. Add LB + agar into a 2L autoclavable glass flask, add 1000 mL dH₂O.4. Cover the glass flask top with aluminum foil. Autoclave at liquid setting for 20 minutes in a basin.5. Let LB/agar solution cool to $\sim 55\text{ }^\circ\text{C}$ under room temperature (you should be able to pick up the jar without a glove), add appropriate antibiotics according to your need, mix well by swirling the flask using hands. Pouring the plates <ol style="list-style-type: none">1. Make sure bench top has wiped down with bleach/EtOH.2. Remove sterile 100 mm Petri dishes from plastic bag (save the bag for storage).3. Pour a thin layer of LB Agar ($\sim 10-15\text{ mL}$) into each plate being careful to not lift the cover off excessively (you should be able to just open up enough to pour).4. Swirl plate to distribute LB/agar on the entire plate.5. Let each plate cool until its solid ($\sim 20\text{ minutes}$) then flip so as to avoid condensation on the agar.6. Put plates back into the bag, store plates at $4\text{ }^\circ\text{C}$.